



Fig. 1. Hordeivirus morphology. Electron micrograph of negatively stained particles of BSMV. The virus particles are ~25 nm in width and vary in length from ~100 to 160 nm.

The genome organization of BSMV is shown in **Fig. 2**. All three RNAs have a 7-methylguanosine cap at the 5' terminus and a conserved 3' region containing an internal poly(A) tail directly following the stop codon of the 3' proximal open reading frame (ORF). Following the poly(A) tail is a 238 nucleotide tRNA-like structure that is capable of binding tyrosine *in vitro*. The  $\alpha$ ,  $\beta$ , and  $\gamma$  genomes are required to establish a systemic infection in plants; the  $\alpha$  and  $\gamma$  genomes are sufficient for viral RNA replication in protoplasts (5). The  $\alpha$  genome encodes a single 130-kDa protein ( $\alpha$ a) that contains both methyl transferase- and nucleotide-binding domains. At least four proteins are encoded by the  $\beta$  genome:  $\beta$ a (23 kDa) is followed by an intergenic region that precedes a series of overlapping genes, designated the triple gene block. The triple gene block encodes proteins of 58 kDa ( $\beta$ b), 14 kDa ( $\beta$ d), and 17 kDa ( $\beta$ c). The  $\beta$ a protein is the BSMV capsid protein, but, despite its role in formation of virus particles, it is not essential for systemic spread (6).  $\beta$ b is known to bind RNA and nucleotide triphosphates *in vitro*. Hence it is speculated that it has a role as an RNA helicase *in vivo*, even though *in vitro* helicase activity has not been associated with the purified protein (7).  $\beta$ d and  $\beta$ c are both hydrophobic proteins, and immunolocalization of  $\beta$ d in infected barley tissue has shown that it