

these cases, the reaction conditions should be calculated as described in Chapters 28, 41, 42, and 48. *Agrobacterium* colonies without an insert should be used as a control.

References

1. Heie, Y., Ohta, S., Komari, T., and Kumashiro, T. (1994) Efficient transformation of rice (*Oryza sativa* L.) mediated by *Agrobacterium* and sequence analysis of the boundaries of the T-DNA. *Plant J.* **6**, 271–282.
2. de Framond, A., Barton, K. A., and Chilton, M.-D. (1983) Mini Ti: a new vector strategy for plant genetic engineering. *Biotechnology* **1**, 262–269.
3. Zambryski, P., Joos, H., Genetello, C., Leemans, J., Van Montagu, M., and Schell, J. (1983) Ti-plasmid vector for the introduction of DNA into plant cells without alteration of their normal regeneration capacity. *EMBO J.* **2**, 2143–2150.
4. Herrera-Estrella, L. and Simpson, J. (1988) Foreign gene expression in plants, in *Plant Molecular Biology: A Practical Approach* (Shaw, C. H., ed.), IRL, Oxford, UK, pp. 131–160.
5. Nagel, R., Elliott, A., Masel, A., Birch, R. G., and Manners, J. M. (1990) Electroporation of binary Ti plasmid vector into *Agrobacterium tumefaciens* and *Agrobacterium rhizogenes*. *FEMS Microbiol. Letts.* **67**, 325–328.
6. Holsters, M., De Waele, D., Depicker, A., Messens, E., Van Montagu, M., and Schell, J. (1978) Transfection and transformation of *Agrobacterium tumefaciens*. *Mol. Gen. Genet.* **163**, 181–187.
7. Schledzewski, K. and Mendel, R. R. (1994) Quantitative transient gene expression: comparison of the promoters for maize polyubiquitin1, rice actin1, maize-derived *Emu* and CaMV 35S in cells of barley, maize and tobacco. *Transgenic Res.* **3**, 249–255.
8. Grant, J. E., Cooper, P. A., McAra, A. E., and Frew, T. J. (1995) Transformation of peas (*Pisum sativum* L.) using immature cotyledons. *Plant Cell Rep.* **15**, 254–258.
9. Bevan, M. W. (1984) Binary *Agrobacterium* vectors for plant transformation. *Nucleic Acids Res.* **12**, 8711–8721.
10. Quemada, H. D., Gonsalves, D., and Slightom, J. L. (1991) Expression of coat protein gene from cucumber mosaic virus strain C in tobacco: protection against infections by CMV strains transmitted mechanically or by aphids. *Phytopathology* **81**, 794–802.
11. Namba, S., Ling, K., Gonsalves, C., Slightom, J. L., and Gonsalves, D. (1992) Protection of transgenic plants expressing the coat protein gene of watermelon mosaic virus II or zucchini yellow mosaic virus against six potyviruses. *Phytopathology* **82**, 940–946.
12. Ling, K., Namba, S., Gonsalves, C., Slightom, J. L., and Gonsalves, D. (1991) Protection against detrimental effects of potyvirus infection in transgenic tobacco plants expressing the papaya ringspot virus coat protein gene. *Biotechnology* **9**, 752–758.
13. Hooykaas, P. J. J. and Schilperoort, R. A. (1992) *Agrobacterium* and plant genetic engineering. *Plant Mol. Biol.* **19**, 15–38.