

## 2.11. Amplification of the Primary Library

1. Packaged and titered primary library.
2. Prepared, appropriate *E. coli* host strains.
3. NZY medium, plates, and top agarose: (see Subheading 2.10., item 1).
4. SM buffer: (see Subheading 2.10., item 5).
5. Chloroform.
6. Dimethylsulfoxide (DMSO).

## 3. Methods

### 3.1. First-Strand Synthesis

The final volume of the first strand synthesis reaction should be 50  $\mu\text{L}$ . Take this into account when determining the volumes necessary.

1. In an RNase-free microcentrifuge tube, add the reagents in order: 5.0  $\mu\text{L}$  10X first-strand buffer, 3.0  $\mu\text{L}$  methyl-nucleotide mixture, 2.0  $\mu\text{L}$  linker-primer (1.5  $\mu\text{g}/\mu\text{L}$ ), X L DEPC-treated water, 40 U ribonuclease inhibitor.
2. Mix the reagents well. Add X  $\mu\text{L}$  of poly(A)<sup>+</sup> mRNA (5  $\mu\text{g}$ ) and gently vortex (see Notes 1 and 2).
3. Allow the mRNA template and linker-primer to anneal for 10 min at room temperature.
4. Add 0.5  $\mu\text{L}$  of [ $\alpha$ -<sup>32</sup>P]-labeled deoxynucleotide (800 Ci/mmol). Do not use [ $\alpha$ -<sup>32</sup>P]dCTP (see Note 3).
5. Add 250 U of reverse transcriptase. The final volume of the reaction should now be 50  $\mu\text{L}$ .
6. Gently mix the sample and briefly spin down the contents in a microcentrifuge.
7. Incubate at 37°C for 1 h.
8. After the 1-h incubation, place on ice.

### 3.2. Second-Strand Synthesis

The final volume of the second strand synthesis reaction should be 200  $\mu\text{L}$ . Take this into account when determining the necessary volumes.

1. To the first-strand reaction (50  $\mu\text{L}$ ), add the following components in order: 20.0  $\mu\text{L}$  10X second-strand buffer, 6.0  $\mu\text{L}$  second-strand dNTP mixture, X  $\mu\text{L}$  sterile distilled water (DEPC-treated water is not required), 4 U *E. coli* RNase H, 100 U *E. coli* DNA polymerase I.
2. The final volume of the reaction should now be 200  $\mu\text{L}$ . Quickly vortex and spin down the reaction in a microcentrifuge. Incubate for 2.5 h at 16°C.
3. After the 2.5-h incubation, place on ice.

### 3.3. Blunting cDNA Termini

1. Add the following reagents to the synthesized cDNA: 23.0  $\mu\text{L}$  blunting dNTP mixture, 2.0  $\mu\text{L}$  cloned *Pfu* DNA polymerase (2.5 U/ $\mu\text{L}$ ).